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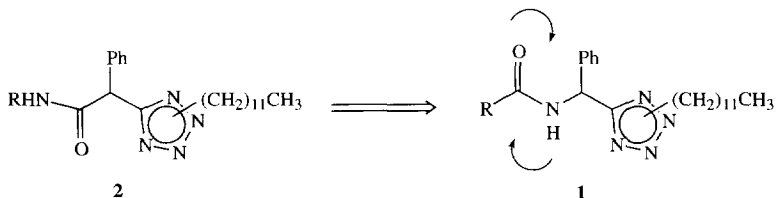
**INHIBITORS OF ACYL-CoA:CHOLESTEROL O-ACYL
TRANSFERASE (ACAT) AS HYPOCHOLESTEROLEMIC AGENTS. 13.
DESIGN, SYNTHESIS AND BIOLOGICAL EVALUATION OF
TETRAZOLE ANILIDES AS POTENT INHIBITORS OF ACAT IN VITRO AND
HYPOCHOLESTEROLEMIC AGENTS IN VIVO.**

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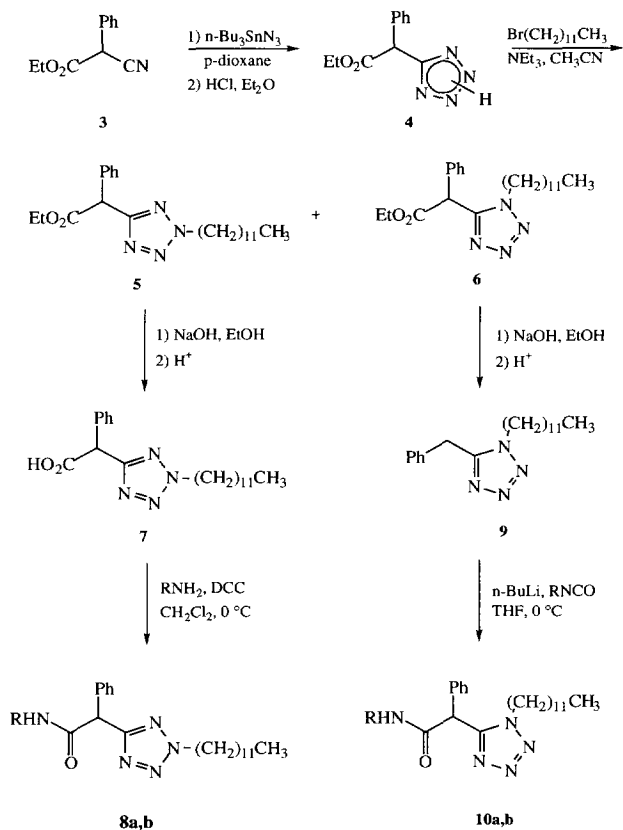
Abstract The syntheses and biological activities for anilides derived from 2-phenyl-2-(dodecyl-2H-tetrazol-5-yl)acetic acid are described. Evidence is provided that one of these compounds, (+)-**8b**, stereoselectively inhibits ACAT *in vitro* and possesses superior efficacy *in vivo* compared to (-)-**8b** or the racemic mixture (\pm)-**8b**.

We have recently reported on a series of benzamide and nicotinamide derivatives (**1**) that potently inhibit ACAT *in vitro* and are efficacious in lowering plasma total cholesterol *in vivo*.¹ Evidence was provided that the (*N*-dodecyltetrazol-5-yl)-benzyl moiety in **1** is necessary for potent inhibition, since replacing this functionality with an oleyl side chain or varying the tetrazole chain length, resulted in a marked reduction in inhibitory activity. In a related study however, oleic acid anilides, substituted with electron donating groups on the 2,6- or 2,4,6-aryl positions, potently inhibit ACAT *in vitro* with IC₅₀'s in the 7 to 700 nM range.² Since fatty acid anilides have been shown to be significantly more potent than the corresponding benzamide isosteres,^{1,3} we sought to improve the *in vitro* activity of the tetrazole derivatives by combining optimal structural features from both series, by replacing the benzamide bond of **1** with 2,6-diisopropyl- and 2,4,6-trimethoxy substituted anilide bioisosteres (**2**). Of the compounds prepared, a trimethoxyphenyl analog of **2** was resolved into individual enantiomers, in order to assess whether the biological activity observed for **2** resided in one particular enantiomeric form. In this paper, we will describe the syntheses and biological results for this novel series of tetrazole amide ACAT inhibitors.



Chemistry: The racemic anilides **8a,b** and **10a,b** were prepared employing the synthetic route illustrated in Scheme 1. The tetrazole intermediate **4** was synthesized in 66% yield by treating (\pm)-ethyl phenylcyanoacetate **3** with *n*-tributyltin azide in refluxing *p*-dioxane, with subsequent cleavage of tributyltin from the tetrazole moiety with ethereal HCl. Alkylation of **4** with 1-bromododecane in refluxing acetonitrile provided a 2.5:1 mixture of regioisomers **5** (52%) and **6** (21%).⁴ This isomeric mixture could be easily separated by silica gel chromatography, or taken on to the next step as a crude mixture. Alternatively, **5** can be prepared by isomerizing the mixture by heating neat in iodododecane at 140 °C.⁵ Saponification of **5** gives the expected carboxylic acid

Scheme 1



derivative **7** (90%). However, **6** decarboxylates to **9** quantitatively upon treatment with sodium hydroxide in ethanol. This proved to be an effective method of separating the 1- from the 2-regioisomer without the use of chromatography, since **9** is insoluble in aqueous base, whereas the sodium salt of **7** is highly soluble. Filtration of **9** followed by acidification of the filtrate, gave **7** exclusively as the 2-regioisomer. Amides **8a,b** were then prepared by coupling an appropriately substituted aniline with **7** in dichloromethane using DCC as the coupling agent (60%). To prepare the 1-regioisomers **10a,b**, the benzylic tetrazole **9** was deprotonated with *n*-butyllithium in THF, with subsequent quenching of the anion with 2,6-diisopropylphenyl- or 2,4,6-trimethoxyphenyl isocyanate (40%).

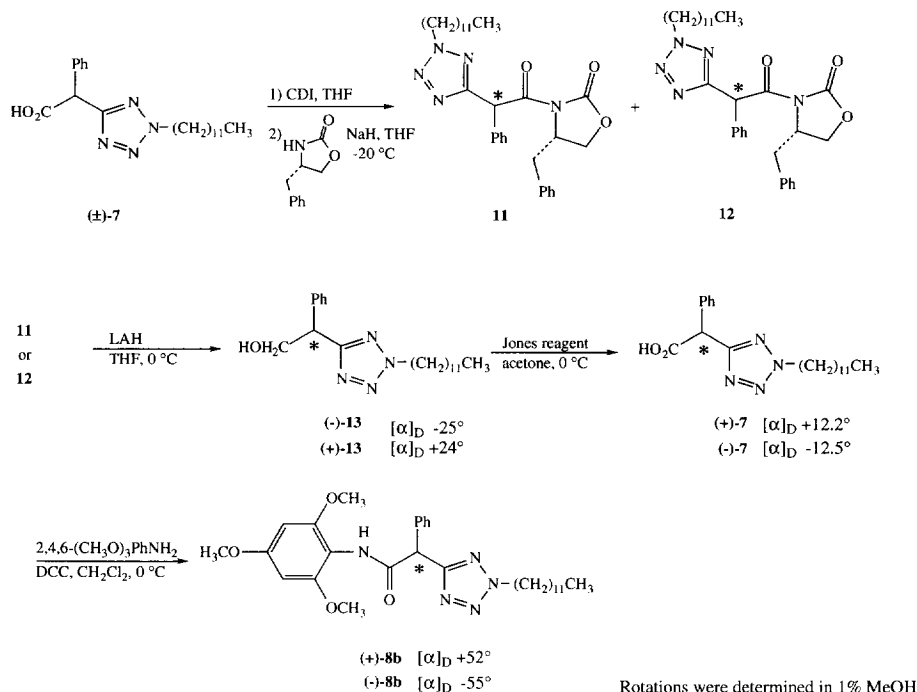
The enantiomers of **8b** were initially isolated by chiral preparative HPLC. The racemate, **8b**, was dissolved in a solution of 80:20 2-propanol:hexane and injected onto a 500 x 20.0 mm Chiralcel OG® preparative column at a flow rate of 8.0 ml/minute. The first enantiomer to elute, (-)-**8b**, was found to be 98% enantiomerically pure ($[\alpha]_D = -58.0^\circ$, $c=1\%$ MeOH) by HPLC. The second enantiomer isolated, (+)-**8b**, was 96.3% pure ($[\alpha]_D = +55.1^\circ$, $c=1\%$ MeOH), but contained 3.7% of (-)-**8b**. After 19 injections, 1.85 grams of (±)-**8b** yielded 708 mg of (-)-**8b** and 727 mg of (+)-**8b**.

With the enantiomers in hand, we performed a solution stability study to determine the racemization potential for the respective isomers at 37 °C. Compounds (+)-**8b** and (-)-**8b** were evaluated at pH's of 1.0, 7.4 and 8.5 over a 25-hour period by chiral HPLC. In an acidic media, no racemization was observed. At a physiological pH of 7.4 however, the half-life of racemization is 56 hours for (+)-**8b** and 62 hours for (-)-**8b**. The rate of racemization is significantly increased (11 and 6 hours respectively) at pH 8.5.

In order to prepare quantities of (+)- and (-)-**8b** sufficient for *in vivo* biological testing, several resolution strategies were examined. Attempt to resolve **8** (Scheme 1) using classical resolution techniques were ineffective. As shown in Scheme 2, however, utilizing Evan's oxazolidone chiral auxiliary,⁶ we were successful in preparing **8b** in either enantiomeric form. Thus, racemic acid **7** was coupled to the sodium salt of (4*S*)-benzyl-2-oxazolidone using CDI in THF to give a 1:1 mixture of diastereomers **11** and **12**. These compounds were separated pure (>99%), but in low yield (35%), using silica gel chromatography (elution with hexane/ THF (5:1) gave **11** and **12** respectively). Reduction of the imide functionality in **12** with LAH in THF provided the resolved alcohol (+)-**13**⁷ (78%), followed by Jones oxidation at 0 °C to the penultimate acid (-)-**7** in 42% yield. Treatment of (-)-**7** with 2,4,6-trimethoxyaniline and DCC in dichloromethane yielded the target compound (50%), (-)-**8b**, in 93% enantiomeric excess.⁸

Identical reaction conditions were used converting **11** to (+)-**8b**.

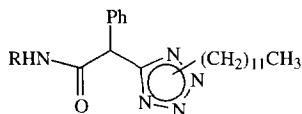
Scheme 2



Results and Discussion: The biological activity associated with the isosteric modification of **1** with 2,6-diisopropyl- and 2,4,6-trimethoxy-substituted anilides is shown in Table I.⁹ The initial compounds prepared for this study, **8a** and **8b**, are considerably more potent than the corresponding benzamide tetrazoles **1** previously reported (compare IC₅₀'s of 10 and 24 nM to 810 and 1700 nM respectively).¹ This enhancement in *in vitro* potency correlates well with the *in vivo* efficacy obtained in the APCC screen, since both **8a** and **8b** significantly lower plasma cholesterol (-46%, -65%) at a dose (3 mg/kg) whereas the corresponding benzamide derivatives are ineffective. Positioning of the tetrazole alkyl chain in these compounds favors the 2-position, since the 1-regioisomers, **10a** and **10b**, were less active than **8a** and **8b** both *in vitro* and *in vivo*. A further assessment of hypocholesterolemic activity for **8a** and **8b** was measured in the CPCC

screen. In this model, **8b** was considerably more efficacious than **8a**, lowering plasma total cholesterol by 59% at 10 mg/kg, whereas the reduction in plasma total cholesterol with **8a** did not achieve statistical significance at doses as high as 30 mg/kg. In this series of biological assays, **8b** also demonstrated superior *in vitro* potency and efficacy *in vivo* compared to CI-976.

TABLE I



Compound ^a	R	Isomer	IC ₅₀ (μM) ^b	APCC (%ChangeTC) ^c		CPCC (%ChangeTC) ^d	
				30 mg/kg	3 mg/kg	10 mg/kg	3 mg/kg
CI-976			0.110	-57*	-15	-17	-12
8a	2,6-diPr ₂ Ph	2	0.010	-49*	-46*	-38 ^e	-
8b	2,4,6-(CH ₃ O) ₃ Ph	2	0.024	-63*	-65*	-59*	-23
8b (+)	2,4,6-(CH ₃ O) ₃ Ph	2	0.014	-68*	-66*	-60*	-55*
8b (-)	2,4,6-(CH ₃ O) ₃ Ph	2	0.330	-64*	-47*	-11	+7
10a	2,6-diPr ₂ Ph	1	0.079	-44*	-	-24 ^e	-
10b	2,4,6-(CH ₃ O) ₃ Ph	1	0.052	-38*	-	-	-

^a Analytical results are within $\pm 0.4\%$ of the theoretical values. ^b ACAT inhibition *in vitro*, liver microsomes isolated from cholesterol-fed rats. Each determination performed in triplicate. See reference 9 for complete protocol. ^c Denotes percent change in total cholesterol in cholic acid (0.5%)-cholesterol (1.5%)-peanut oil (5.5%)-fed rats administered a single dose of test compound. See reference 9 for the complete protocol. ^d Denotes percent change in total cholesterol using the diet as in footnote c, dosing over a period of 7 days. See reference 9 for the complete protocol. ^e Denotes the compound was tested at 30 mg/kg. * Denotes significantly different from control, $p < 0.05$ using analysis of variance followed by Fisher's multiple range test.

Due to the excellent activity of **8b**, we resolved the racemic mixture into its individual enantiomers. The *in vitro* evaluation of these isomers suggested that (+)-**8b**, may stereoselectively inhibit ACAT, since it is nearly twice as potent as the racemate and greater than 20 fold more potent than (-)-**8b** (14 nM vs 330 nM). It is noted that the 2% of (+)-**8b** present in (-)-**8b**, could account for its residual potency (IC₅₀ = 330 nM). This stereoselectivity is in agreement with a communication recently published by McCarthy et al., demonstrating that (*S*)-*N*-(2,4-bis(methylthio)-methylpyridin-3-yl)-2-(hexylthio)-decanoic acid amide is 7-fold more potent than the corresponding *R*-stereoisomer (IC₅₀ = 22 and 160 nM respectively).¹⁰ Although (+)-**8b** is the preferred stereoisomer *in vitro*, it is equiefficacious with (-)-**8b** and the racemate when administered in a single dose to cholesterol-fed rats. Based on these results, we assumed the asymmetric center epimerizes *in vivo*. However, the enantiomers could be differentiated in the chronic *in vivo* assay. In this assay, (-)-**8b** was shown to be inactive at 10 and 3 mg/kg whereas

(+)-**8b** lowers plasma total cholesterol by 60 and 55% respectively. The racemate was also less effective than (+)-**8b** in this study.

In summary, we have identified 2-phenyl-2-(dodecyl-2H-tetrazol-5-yl)acetic acid as a fatty acid mimetic, that when coupled with 2,6-diisopropyl- and 2,4,6-trimethoxyaniline, yields novel bioisosteric replacements for **1** that are extremely potent *in vitro* and efficacious *in vivo*. Of the compounds prepared, **8b**, is significantly more potent and efficacious than the fatty acid amides previously reported,² including CI-976. We have demonstrated that the majority of the ACAT inhibition observed for **8b** resides in the (+)-enantiomer, (+)-**8b**, since the (-)-enantiomer (-)-**8b**, was significantly less potent *in vitro* and ineffective when administered to hypercholesterolemic rats. Further extensions of this study will be the topic of future communications from this laboratory.

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References:

1. O'Brien, P. M.; Sliskovic, D. R.; Anderson, M. K.; Bousley, R. F.; Krause, B. R.; Stanfield, R. L. Previous paper in this series.
2. Roth, B. D.; Blankley, C. J.; Hoefle, M. L.; Holmes, A.; Roark, W. H.; Trivedi, B. K.; Essenburg, A. D.; Kieft, K. A.; Krause, B. R.; Stanfield, R. L. *J. Med. Chem.* **1992**, *35*, 1609.
3. Roark, W. H.; Roth, B. D.; Holmes, A.; Trivedi, B. K.; Kieft, K. A.; Essenburg, A. D.; Krause, B. R.; Stanfield, R. L. *J. Med. Chem.* **1993**, *36*, 1662.
4. The isomeric mixture was evaluated by ¹HNMR prior to chromatography. Assignments were made based on the chemical shift of the methylene protons attached to the tetrazole ring (see: Scott, F. L.; Tobin, J. C. *J. Chem. Soc. C* **1971**, 703).
5. Isida, T.; Kozima, K.; Nabika, K.; Sisido, K. *J. Org. Chem.* **1971**, *36*, 3807.
6. Evans, D. A.; Britton, T. C.; Dorow, R. L.; Dellaria, J. F. *J. Amer. Chem. Soc.* **1986**, *108*, 6395.
7. The enantiomeric purity was assessed by ¹HNMR using Mosher's acid chloride. Both (-)-**13** and (+)-**13** were considered isomerically pure by this method.
8. The enantiomeric excess was determined using a Chiracel OG 10 micron (4.6 x 250mm) column with a mobile phase of 80:20 hexane:IPA and a flow rate of 1ml/min. Retention time for **8b**(-) is 15.2 min and **8b**(+) is 17.8 min.
9. Krause, B. R.; Black, A.; Bousley, R.; Essenburg, A. D.; Cornicelli, J.; Holmes, A.; Homan, R.; Kieft, K. A.; Sekerke, C.; Shaw-Hes, M. K.; Stanfield, R. L.; Trivedi, B. K.; Woolf, T. *J. Pharm. Exp. Ther.* **1993**, *267*, 734.
10. McCarthy, P. A.; Hamanaka, E. S.; Marzetta, C. A.; Bamberger, M. J.; Gaynor, B. J.; Chang, G.; Kelly, S. E.; Inskeep, P. B.; Mayne, J. T.; Beyer, T. A.; Walker, F. J.; Goldberg, D. I.; Savoy, Y. E.; Davis, K. M.; Diaz, C. L.; Freeman, A. M.; Johnson, D. A.; Lacour, T. G.; Long, C. A.; Maloney, M. E.; Martingano, R. J.; Pettini, J. L.; Sand, T. M.; Wint, L. T. *J. Med. Chem.* **1994**, *37*, 1252.

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